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Sarcoglane, A New Cytotoxic Diterpene from Sarcophyton glaucum

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Abstract: Sarcoglane (1) a new tricyclo[7,5,0^{10,14}]tetradecane diterpenoid has been isolated from the Indo-Pacific soft coral Sarcophyton glaucum. The structure of compound 1, which is cytotoxic to fertilized sea urchin eggs, was defined by spectral methods and mainly 1D and 2D NMR experiments. Sarcoglane possesses an unprecedented tricyclic carbon skeleton, biogenetically, closely related to the cembrane and xeniaphyllane diterpenoids. Copyright © 1996 Elsevier Science Ltd

The interest in the chemistry of soft coral metabolites brought us to reinvestigate Sarcophyton glaucum. Since the isolation of sarcophine, deoxysarcophine and several other cembranoid alcohols from the Red Sea S.glaucum² a whole series of other cembranoids have been isolated from S.glaucum collected in other seas³. Investigation of a Indo-Pacific S.glaucum collected in Sodwana Bay, Durban, South Africa during January 1995 (IBS-72) afforded in addition to common cembrane and sterol soft coral metabolites¹ a new cytotoxic, to fertilized sea urchin eggs⁴, diterpenoid designated sarcoglane (1). Compound 1 was obtained from the EtOAc extract of the soft coral by solvent partitioning followed by silica gel chromatography (hexane-EtOAc). Sarcoglane m.p.138 0 , $\left[\alpha\right]_{D}$ 110 0 , v_{max} 3540 (OH), 1690 (CO) cm $^{-1}$ had a molecular formula of C20H32O2 as revealed by both EIMS and NMR data (Table 1). The IR and NMR spectra implied the presence of a ketone, a trisubstituted double bond and one hydroxyl group, thus, according to the five degrees of unsaturation, 1 had to be tricyclic. Interpretation of the 1D and 2D NMR spectra (COSY, TOCSY, HMQC and HMBC) suggested two moieties: -CH2CH2CH(iPr)CHCHCH- and -CH2CH2CH=CMeCH2CO- (Table 1, C-1 to-13 and C-2 to-7) which together with two tertiary methyl groups (\$C-CH3 and HO-C-CH3) accounted for all 20 carbon and 32 hydrogen atoms of 1. Connection of the partial structures of 1 based on CH-correlations (HMBC spectrum, Table 1) established a tricyclic structure for sarcoglane. Most significant were the strong 2J and 3J CH-couplings of the five methyls and methines with their neighbors in the molecule. Consequently, a 8-hydroxy-11-isopropyl-4,8,14trimethyltricyclo-[7,5,0^{10,14}] tetradec-4-ene-2-one structure was unambiguously determined for 1.

With the planar structure of 1 in hand, the relative stereochemistry was probed by assignment of H-H coupling constants and by a series of NOE enhancements as summarized in Table 1. The NOE's between Me-20(β) and H-1 and H-10 established the β -orientation of the latter two protons and, hence, also the cis junction between the cyclobutane and cyclopentane rings. A NOE between H-1 β and Me-16 determined the

β-conformation of the latter methyl group. As in caryophyllene⁵ and the xeniaphyllanes⁶ the cyclononene ring may adopt several different conformations. In case of 1 the major conformation seem to be the one with the Me-15β orientation based on a NOE between the latter methyl and Me-16β (a NOE which also determined the 8S* configuration). The carbon chemical shift of Me-15 (δc 19.8ppm - due to a γ-effect with CH₂-6) established the E-configuration of the double bond, thus, H-5 has to be α. Finally, NOE enhancements between H-5α and H-9 and H-11 determined the relative configuration of C-9 and C-11, and the trans ring junction between the four and nine membered rings. Further support for the cyclobutane ring came from the one bond CH-coupling constants of 132, 132 and 135 Hz for C-1, 9 and 10, respectively, as well as from characteristic MS-fragmentations of this ring⁸. Biogenetically, it is difficult to make the distinction whether sarcoglane is obtained from geranyl geraniol via a cembrane or xeniaphyllane precursor,

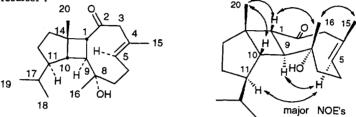


TABLE 1. NMR Data of sarcoglane (1) (500 MHz, in CDCl.)

							•	. ,	•		3′			
ł] #	δ_{c}	$\delta_{_{_{\rm H}}}$	J(Hz)	COSY	HMBC	NOE	Н#	$\delta_{_{\rm C}}$	$\delta_{_{\rm H}}$	J(Hz)	COSY	HMBC	NOE
					TOCSY (C to H#)						TOCSY	(C to H#)	
1	l	54.1d	2.45d	8.5	9	13′,20		11	55.2d	1.50m		10,12,12'	17,18,19	
3	?	208.6s 53.7t	2.65d	15.4	3′	1,3,3′		12	29.5t	1.64m 2.20m		11,12',13,13 11,12,13,13'		
4	ļ	126.3s	3.15brd	15.4	3,5	3,3′,15		13	33.3t	1.19m 2.30m		12,12',13' 12,12',13	12,20	
5	5	134.9d 25.3t	4.79d 2.08brd	11.7 13.8	3,6,15 6',5,7	3,15	9,11	14	44.3s	-		12,12,13	1,10,13,20	
·	,	23.31	2.28dt	4.0,13.8	6,7,7'	,			19.8q 29.3q	1.85s 1.15s		5		3,16 1,6,15
7	7	46.8t	1.65dt 1.95dt	4.0,13.8 14.0,4.0	6,6′,7′ 6′,7	16		17	30.6d	1.38sep	6.5	18,19	18,19	
8		75.5s	-	•	•	1,7,9,10,1	6		20.5q 20.6q	1.10s 0.85d	6.5	17 17	17,19 18	1,10
9 1		56.3d 49.4d	2.02dd 1.98m	8.5,7.5	1,10 9,11	1,7,10,16 9,19,20			21.8q	0.90d	6.5	17	1	

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- EIMS m/z(%): 304(M⁺,15), 286(M-H₂O,10), 261(M-Pr,12), 181(C₁₁H₁₇O₂, the cyclononane ring, 46), 123(C9H₁₅, the cyclopentane ring, 80), 81(124-Pr, 100).
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